



PROCEEDINGS OF THE 12th WORLD RABBIT CONGRESS

Nantes (France) - November 3-5, 2021 ISSN 2308-1910

This communication was accepted by the scientific committee of the Congress

but was not presented during the Congress itself, neither face-to-face nor remotely via Internet.

MOLECULAR IDENTIFICATION OF PATHOGENS IN THE ENTERIC SYNDROME IN RABBITS FROM MEXICO

Bautista-Gómez L.G.^{1*}, Martínez-Castañeda J.S.², Reynoso-Utrera E.¹, Jiménez- Ramos A.¹, López Aguado-Almazán G.¹, Trejo-Huitrón G.¹, Rodríguez-Villavicencio V.¹, Aguilar-Gutiérrez M.³

¹Centro Universitario UAEM Amecameca, Universidad Autónoma del Estado de México, Km 2.5 Carretera Amecameca-Ayapango, C.P. 56900 Amecameca, Estado de México, México

²Centro de Investigación y Estudios Avanzados en Salud Animal, Facultad de Medicina Veterinaria y Zootecnia, Universidad Autónoma del Estado de México. Carretera de Cuota Toluc a-Atlacomulco km 15.5, C.P. 50200, Toluca, Estado de México, México ³Unidad de alta especialización en pediatría. Clínica Médica, Nuestra Señora Del Rosario, Amecameca,

CP. 56900, Estado de México, México.

*Corresponding author: lin_bag@yahoo.com.mx

ABSTRACT

In Rabbits the enteropathies are difficult to diagnose; their etiology involves different pathogens that act concomitantly, causing damage to the intestine. The aim of the present study was identification by molecular techniques of the pathogens present in enteric rabbits in Mexico. From May 2014 to December 2019, we screened 228 samples of the intestinal content of rabbits having a clinical history of enteric disease. Out of the samples analyzed, Eimeria spp. were found in 63.59%, of this E. Vejdovski was identify in the 85% of the one sample screened by specific primers. Followed by Bacteria (49.15%) of this Escherichia coli was present in the 33.23%, which were identified as enteropathogenic E. coli (EPEC), Viruses was present in the 14.47% of the analyzed samples.

Key words: Rabbits, Enteropathies, Pathogens, Mexico.

INTRODUCTION

Enteric diseases have an important role in breeding farms, as they cause serious economic losses due to mortality, growth depression and decline in the conversion rate [Lavazza et.al. 2008; Papp et. al. 2013]. The observed symptoms include anorexia, polydipsia, weakness, abdominal distension, profuse diarrhea, dehydration, hypothermia and death. Enteropathies are recognized as a problem for domestic rabbits, being their etiology and pathogenesis poorly understood. In commercial rabbit production, enteropathies are hard to diagnose; they can have a multifactorial etiology.

Objectives : The aim of this study was to isolate pathogens from enteric disease in rabbits from Mexico.

MATERIALS AND METHODS

Animals and experimental design

Two hundred twenty-eight rabbits, with an enteric clinical profile were obtained from several rabbit meat production from the southeastern part of the State of Mexico, Mexico. The animals exhibited an enteric clinical profile that included depression, anorexia, dehydration, abdominal distension, liquid-to-mucoid diarrhea. The affected animals were young rabbits of 40-60 days old, they were humanely slaughtered according to the Official Mexican Standard NOM-033-SAG/ZOO-2014, and examined *post mortem*. Bacteriological and coproparasitoscopic examination was performed from intestinal contents. This study was authorized by the Bioethics Committee of the CU Amecameca (CBE/06/2013).

Pathogens Identification

Intestinal content samples were assessed by a coproparasitologic study by means of the McMaster technique to detect the qualitatively and quantitatively Eimeria spp. oocysts present. Of the positive samples from the coproparasitoscopic examination, we proceeded to extract total DNA directly from 1 g of faeces, using the ZR faecal Microbe DNA Zymo Research®, following the manufacturer's instructions. The obtained DNA was visualised by electrophoresis on agarose gel 1%. For molecular identification, we used the primers ITS1F (5'GGGAAGTTGCGTAAATAGA 3') and ITS1R (5'CTGCGTCCTTCATCGAT 3') reported by Oliveira et al. (2011) that amplified a fragment of 400–600 bp for the, region of the eleven Eimeria species that affect rabbits. The amplification conditions were; Initial denaturalization at 96 °C for 10 minutes, 30 cycles consisting of 96 °C for 45 seconds, 61 °C for 45 seconds at and 72 °C for 1 minute at with a final extension step of 72 °C for 5 minutes. The amplified products are separated by electrophoresis on agarose 3% gels and stained with ethidium bromide.

The bacteriological analyses involved primary isolation in blood agar; identification was performed by colonial morphology and Gram staining. Gram negative bacteria were cultured in the following selective media: MacConkey, Salmonella-Shigella and brilliant green agar at 37 °C incubation temperature and aerobiosis for 24-48 h.

The virological examination consisted of the detection of the VP4 and VP7 genes of rotavirus through the reverse polymerase chain reaction (RT-PCR), for norovirus, we amplified a 350 bp fragment of ORF1, that synthesizes a protein RDRP (RNA dependent of RNA polymerase), and for Astrovirus a fragment of 409bp of ORF1b was amplified. From the total RNA extracted from the sample. Primers reported by Gentsch et al., (1992) were used for the amplification of an 876 bp fragment that encodes the VP4 protein (Con 3 [nt 11–32] F 5' TGGCTTCGCCATTTTATAGACA 3' and Con 2 [nt 887– 868] R 5' ATTTCGGACCATTTATAACC 3'). For VP7 amplification, the primers reported by Gouvea et al., (1990) were used, (Beg 9 [nt 1-28] F 5' GGCTTTAAAAGAGAGAAATTTCCGTCTGG 3' and End 9 [nt 1062-1036] R 5' GGTCACATCATACAATTCTAATCTAAG 3'). The primers TGACAATGTAATCATCACCATA y NoVRev TGACGATTTCATCATCACCATA NoVFor (Caddy, et. Al. 2015), was used for Norovirus and panASTVFor2 5'GARTTYGATTGGRCKAGKTAYGA3' and panASTVRev 3'GGYTTKACCCACATNCCRAA5' (Chu et al., 2008). For Astrovirus. Viral RNA was obtained from both tissue and intestinal contents using the GeneJET Viral DNA and RNA Purification kit (Thermo ScientificTM), according to the manufacturer's instructions. Once the viral RNA was obtained, a single-step RT-PCR was performed using the commercial kit SuperScript® III One Step RT-PCR with Platinum® Taq (InvitrogenTM). PCR products were visualized on 2% agarose gels stained with ethidium bromide.

RESULTS AND DISCUSSION

Concurrent identification	Frequency	Percentage
		0
Eimeria spp.	145	63.59
Bacteria	112	49.15
Virus	33	14.47
Eimeria spp. + Bacteria	91	39.91
Eimeria spp. + Virus	12	5.26
Bacteria + Virus	9	3.94
Eimeria spp. + Bacteria + Virus	12	5.26
Astrovirus	12	5.26
Rotavirus	5	2.19
Norovirus	16	7.01

Table 1. Concomitant pathogens found in rabbit intestine samples from Mexico

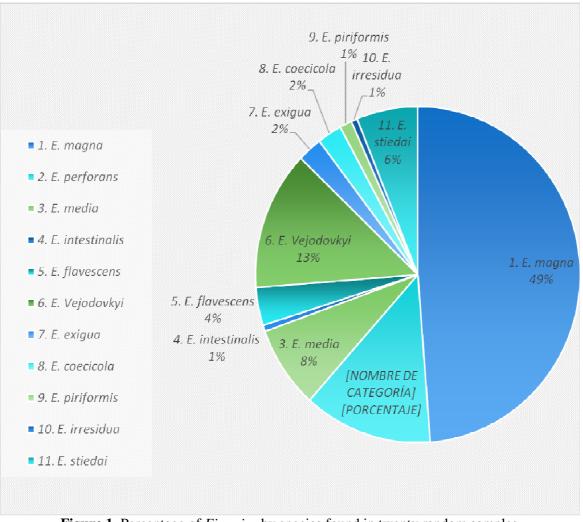


Figure 1. Percentage of Eimerias by species found in twenty random samples

Out of 228 intestine content samples, (100.00%) werepositive for at least one identification and 124 (54.38%) were in concurrence, including: 112 (49.12%) with two identifications and 12 (5.26%) with three identifications. Rotavirus, Norovirus and Astrovirus was identified for the first time in rabbits from Mexico in 33 samples (14.47%), of which 6 samples (2.63%) showed the identification of concomitant infection by Rotavirus, Astrovirus and Norovirus.

Enteric diseases causes 30-80% of deaths in farm animals from 6 to14 weeks old, across the world, by this have an important role in rabbit production, as they can lead to severe economic losses due to mortality, growth depression and reduced feed conversion rate. Enteric syndrome is one of the most important diseases in rabbits, especially in relation to its productive and economic impact. Among the different pathogens that can be found in rabbits with enteric disease, viruses seem to have an important but not definitive role. (Lavazza and Capucci, 2008). Bacteriological and coproparasitoscopic tests identified the presence of potential pathogens in rabbits (Licois, 2004); E. coli in three samples and Eimeria spp.,

Studies in other countries have reported the presence of eleven Eimeria species that affect rabbits in up to 70% of enteropathies, and, according to their level of pathogenicity, can cause reduced growth rate and feed conversion, and increased mortality.

CONCLUSIONS

The presence of different pathogens in more than half of the intestinal content samples of rabbits with enteric symptoms indicates the need to carry out a greater number of studies to determine if these pathogens in isolation cause enteric profiles distinct than when they are found concomitantly.

ACKNOWLEDGEMENTS

To the Autonomous University of the State of Mexico (UAEMex) for funding the present study

REFERENCES

- Caddy SL, de Rougemont A, Emmott E, et al. Evidence for Human Norovirus Infection of Dogs in the United Kingdom. J Clin Microbiol 2015;53:1873–1883.
- Lavazza A, Cerioli M, Martella V, Tittarelli C, Grilli G, Brivio R, Buonavoglia C. 2008. Rotavirus in diarrheic rabbits: prevalence and characterization of strains in Italian Farms. *In Proc: 9th World Rabbit Congress, 10-13 June, 2008. Verona, Italy. 288: 993-998.*
- Lavazza, A., Capucci, L., 2008. Viral infection of rabbits. In Proc: 9th World Rabbit Congress, 10-13 June, 2008. Verona, Italy. 879-883.
- Licois, D., 2004. Domestic rabbit enteropathies. In: Proc. 8th World Rabbit Congress. September, 2004. Puebla, Mexico, 385-403.
- Mogollón JD, Peⁿa N, De Jimeno MC, Rodríguez G. Complejo enteritis mucoide del conejo. I. Descripción de casos de ocurrencianatural. *Revista ICA Bogotá. 1981; XVI:91---6.*
- Oliveira U, Fraga J, Licois D, Pakandl M, Gruber A. Development of molecular assays for the identification of the 11 Eimeria species of the domestic rabbit (Oryctolagus cuniculus). *Vet Parasitol.*2011;176:275--280.
- Papp H, László B, Jakab F, Ganesh B, De Grazia S, Matthijnssens J, Ciarlet M, Martella V, Bányai K. 2013. Review of group A rotavirus strains reported in swine and cattle. *Vet. Microbiol. 165: 190-199.*
- Percy DH, Muckle CA, Hampson RJ, Brash ML. The enteritis complex in domestic rabbits: a field study. Can Vet J. 1993;34:95-102.