IN VITRO CAECAL GAS PRODUCTION AND DRY MATTER DEGRADABILITY OF SOME BROWSE LEAVES IN PRESENCE OF ENZYMES FROM ANAEROBIC BACTERIUM IN NZW RABBITS

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ABSTRACT

This work was conducted to assess the nutritive value of four browse leaves (Acacia saligna, Atriplex halimus, Lucena leucocephala, and Eucalyptus camaldulensis) grown in the northern region of Egypt using the ceacum content obtained directly after slaughtering of 12 New Zealand White rabbits. ZADO®, enzymic preparation containing cellulases, xylanases, α -amylase and proteases from an anaerobic bacterium (patent No. 22155 of Egypt) was added at 0, 2 or 5 mg/ml of buffered caecal content. In vitro caecal gas production after 24 h of incubations (IVGP₂₄) and dry matter degradability (IVDMD) were determined. Crude protein content of browse leaves ranged from 105 g/kg DM (A. halimus) to 221 g/kg DM (L. leucocephala). The highest ether extract (68 g/kg) was in L. leucocephala, while this species had the lowest neutral detergent fibre (NDF), acid detergent fibre (ADF), and acid detergent lignin content. E. camaldulensis had the highest NDF, ADF and lignin. Total extractable phenolics (TEP), saponins (SAP) and alkaloids (ALK) contents were low in L. leucocephala and high in A. halimus and E. camaldulensis. The IVGP₂₄, and IVDMD varied (P<0.001) among leaves, with the highest (P<0.001) value in A. halimus, intermediate in A. saligna and E. camaldulensis, and the lowest (P<0.001) in L. leucocephala. When comparing all leaves species, addition of ZADO[®] improved (P<0.05) caecal IVGP₂₄, and DMD in all leaves without a significant difference (P>0.05) between the two ZADO[®] levels (2 or 5 mg ZADO[®]/ml). ZADO[®] increased the rate of gas production during the first 6 h of incubations. Our results suggest a positive effect of ZADO[®] addition on browse leaves degradation in rabbits. ZADO[®] could have a positive influence on the caecal microbial activity and nutrient digestion, as well as the degradation of secondary compounds of the browse leaves.

Key words: Gas production, Dry matter degradability, ZADO[®], Ceacal activity, Probiotics, Rabbits.

INTRODUCTION

Tree and shrub leaves are important sources of forage for small ruminants (Salem et al., 2006) in the semi-arid conditions of northern Egypt, but the use of locally available foliage in the nutrition of the rabbit is a subject relatively unexplored in tropical countries. This is in spite of the fact that tropical foliages are locally available and at low cost, which is in contrast with the scarcity and often high cost of commercial feeds. However, most tropical browse species contain substantial amounts of phenolic compounds, mainly tannins (Makkar et al., 1995; Salem, 2005), as well as other secondary compounds such are phenolics, saponins and alkaloids, lectins, polypeptides, and essential oils (Salem et al., 2006, 2007), which can reduce their nutritional value, as most of tannins bind to feed proteins thereby making them unavailable to caecal and ruminal microorganisms. Use of tree and shrub leaves by herbivores may be restricted by negative effects of secondary compounds on digestion (Provenza, 1995; Salem, 2005; Salem et al., 2006, 2007). ZADO[®], as a product containing cellulases, xylanases, α -amylase and proteases from an anaerobic bacterium, showed a positive effect on ruminant performance and nutrient utilization of low quality forages in vivo (Gado, 1997) and in vitro (Gado et al., 2007). Probiotics are food ingredients which are supposed to stimulate the activity and potentially alter the composition of the gut flora by providing energy to selected species of the microbial community (Guerra et al., 2007). Studies on the effect of probiotics for improving the nutritive utilization of tree and shrubs leaves in rabbits are limited. Therefore, the objective of the current experiment was to evaluate the utilization of four browse leaves using the ceacal contents of rabbits in the presence of ZADO[®] which is a mixture of cellulases, xylanases, α -amylase and proteases from an anaerobic bacterium.

MATERIALS AND METHODS

Samples of leaves of the browse species *Acacia saligna*, *Atriplex halimus*, *Leucaena leucocephala*, and *Eucalyptus camaldulensis* were randomly and manually harvested from different parts of both young and mature leaves. Leaf samples were dried at 40°C for 72 h in a forced air oven in triplicate to constant weight, ground in a hammer mill to pass 1 mm sieve, and stored in plastic bags for subsequent determination of chemical components, secondary compounds and *in vitro* fermentation using the caecal contents from NZW rabbits. Methods of chemical analyses, i.e. ash, N, neutral detergent fibre (NDF), acid detergent fibre (ADF) and acid detergent lignin, as well as the secondary compounds, i.e. total extractable phenolics (TEP), saponins (SAP), alkaloids (ALK), and aqueous fraction (AF) were as previously described (Salem *et al.*, 2006).

In vitro caecal gas production (IVGP) was determined according to Menke et al. (1979). Approximately 500 mg of dry matter (DM) of each sample were placed in 60 ml syringes with 0, 2, or 5 mg/ml of caecal buffered liquor of ZADO[®], containing cellulase (7.1 U/g), xylanase (2.3 U/g), α-amylase (61.5 U/g) and proteases (29.2 U/g). ZADO® is the patent No. 22155 of Egypt (Molecular Biology Laboratory of the Ain Shams University in Cairo), prepared according to the procedure of Gado (1997). The piston of each syringe was closed with a hard rubber to prevent inflow of water into the syringe during incubation. The experiments were carried out using 12 NZW rabbits. Rabbits were about 14 weeks of age and 2235 g of LW. Rabbits were fed a concentrate mixture (Table 1). Caecal contents were collected directly after slaughtering of rabbits in three runs (four rabbits for each). Caecal contents were combined, homogenized and kept at 39°C in a water bath, flushed with CO₂ before use, and diluted (1:4 w/v) with the culture medium of Makkar et al. (1995) containing bicarbonate buffer, macromineral, micro-mineral, resazurine and reducing solutions. Caecal buffered liquor (30 ml) was pipetted into each syringe and syringes were immediately placed into a water bath at 39°C. Gas volumes were recorded at 2, 4, 6, 10, 12, and 24 h of the incubation. Blank syringes were incubated in each run. At the end of the incubation (24 h), content of each syringe was centrifuged at 15,000 g for 20 min at 4°C. The residual moisture was removed by drying the tubes overnight at 60°C, and then tubes were weighed and dry matter degradability (IVDMD) calculated from differences between initial and residue weights, minus blank tubes. The study was completed at the laboratory of Animal Production, Alexandria University.

Rate of gas production (RGP) and gas yields (GY) were calculated as follow: RGP₄ h (ml/g DM/h) = (volume of gas produced at 2 h)/4 × sample weight (g). Gas yields (GY₂₄ h) were calculated as the volume of gas produced after 24 h (ml gas/g DM) of incubation divided by the amount of substrate apparently degraded. Gas production readings data (ml/0.5g DM) were fitted using the NLIN option of SAS (1999) to the model of France et al. (2000) as: $G = b \times (1 - e^{-k(t-L)})$, where: 'G' was the volume of gas production (ml/0.5 g DM); 'k' was the rate of gas production (per h) from the slowly fermentable feed fraction 'b', and 'L' was the discrete lag time prior to gas being produced. Data were analyzed as a 4 x 3 factorial experiment (4 browse leaves species × 3 treatments: 0, 2 and 5mg of ZADO[®]/ml) using the 'GLM' option of SAS (1999) with methods of Steel and Torrie (1980), to determine differences due to tree species and ZADO[®]. In the case of significant differences, Duncan (1995) multiple-range test was used to separate means within species.

RESULTS AND DISCUSSION

Crude protein content of the browse leaves ranged from 105 to 221 g/kg DM. L. leucocephala had the lowest NDF, ADF and lignin contents, E. camaldulensis had the highest, and A. saligna and A. halimus the intermediate. Secondary compounds, i.e. total extractable phenolics (TEP), saponins (SAP) and alkaloids (ALK)) were at low concentration in L. leucocephala (36, 3 and 0.2 g/kg DM, respectively) and at high concentration in A. halimus (113, 124 and 2.3 g/kg DM, respectively) as well as in E. camaldulensis (102, 15 and 5 g/kg DM, respectively). Aqueous fraction (AF) of lectins, polypeptides and starch, was higher in A. halimus than in the other browse species. Tannins (TEP) concentrations were higher than 50 g/kg of DM in all leaves except those of L. leucocephala (Table 1). However, high levels of secondary compounds may reduce the feed intake, impair the nutrient digestibility or even be toxic in rabbits (Sreekanth et al., 2006). Some variation among different browse leaves could be due to genotypic characteristics in relation to the type of secondary compounds (Salem et al., 2006). In Figure 1 and Table 2, caecal fermentation parameters (i.e. IVGP₂₄ and its parameters B, k, L; and IVDMD) demonstrate differences in nutritional value of the four shrub species that are closely related to their chemical composition (Salem, 2005). The high IVGP₂₄ and IVDMD in A. halimus suggest a higher extent of fermentation in the first 24 h of fermentation in comparison with the other leaves, especially with L. *leucocephala*. Differences in IVGP₂₄ and IVDMD among the leaves could be due to the different content and nature, of their fibre (Rubanza et al., 2003) and secondary compounds (Salem et al., 2007). Indeed, the higher fibre level, as well as high (P<0.05) levels of secondary compounds in *L. leucocephala* (Salem *et al.*, 2006), are almost certainly responsible for reduced gas production in comparison with the other foliages.

	A. saligna	A. halimus	L. leucocephala	E. camaldulensis	Concentrate ^a					
Chemical composition	b									
OM	902	798	901	945	903					
СР	149	105	221	154	207					
EE	39	14	68	41	62					
CF	239	324	203	98	97					
NDF	389	544	372	615	350					
ADF	302	339	208	542	148					
lignin	134	101	121	192	30					
Secondary compounds	c									
TEP	61	112.9	35.74	102.31	ND^d					
SAP	23.5	123.8	3.14	14.58	ND					
ALK	3.19	2.3	0.2	5.02	ND					
AF	68	47.5	7.28	2.4	ND					

Table 1: Nutrient and secondary compound levels (g/kg DM) of browse leaves and also the commercial concentrate fed to NZW rabbit during the experiment

^aCommercial concentrate used in NZW rabbit feeding during the *in vitro* experiment; ^bOM, organic matter; CP, crude protein; EE, ether extract; NDF, neutral detergent fibre; ADF, acid detergent fibre; lignin, acid detergent lignin; ^cTEP, total extractable phenolic compounds; SAP, saponins; ALK, alkaloids; AF, aqueous fraction (lectins, polypeptides, starch; Cowan, 1999); ^dbelow 0.01 g/kg DM



Figure 1: *In vitro* caecal dry matter degradability (IVDMD) of the four shrub species in the presence of 0, 2, or 5 mg of ZADO^{®1}/ml of caecal buffered liquor after 24h of incubation in NZW rabbit. ¹ZADO[®] is a mixture of cellulases, xylanases, α -amylase and proteases from an anaerobic bacterium

Increased IVGP₂₄ and IVDMD in *E. camaldulensis* than in *L. leucocephala* may be due to its lower condensed tannins concentration in leaves. However, differences in degradability among browses could be also due to the different extent of lignification of NDF (Van Soest, 1994). Salem (2005) reported a similar relationship between $IVGP_{24}$ and NDF using the rumen fluid of sheep, cattle and buffalo. Low nutritive value of L. leucocephala and E. camaldulensis could be due to its NDF binding with polyphenolics (Ndlovu and Nherera, 1997). Probiotics are live bacteria or enzymes mixtures, used as feed supplements (Yoon and Stern, 1995). The original concept of feeding live microbes to livestock was based primarily on potentially beneficial effects, including improved establishment of beneficial gut microflora (Fuller, 1999). Addition of low levels of the enzymes mixture from an anaerobic bacterium (ZADO[®]) improved (P<0.001) parameters of the caecal microbial activity (IVGP₂₄ and IVDMD) in all cultures supplied with browse leaves. The improvement varied between the browse leaves and was more pronounced at higher addition of ZADO[®]. Presumably, this could be due to the beneficial effect of ZADO[®] on fibre hydrolysis (Gado et al., 2007; Juskiewicz et al., 2007). Addition of the exogenous fibrolytic enzyme mixture such ZADO[®] improved in vitro rumen fermentation activity and cell wall digestibility of alfalfa stems (Colombatto et al., 2007). However, the mode of action of live microbials in the rumen and the caecum is not completely understood. The presence of lactate-producing bacteria is thought to help the caecal and ruminal microflora to adapt to the presence of lactic acid (Beauchemin et al., 2003; Guerra et al., 2007; Marinho et al., 2007). Beauchemin et al. (2003) reported that the supplementation of the diet with E. faecium increased (P < 0.05) the proportion of propionate and decreased (P<0.10) proportion of butyrate in ruminal fluid compared with the control. This may be explained by stimulating of lactic acid-utilizing bacteria, which produce propionate.

Table 2: *In vitro* caecal gas production IVGP₂₄ (gas production after 24 h of incubation, in ml/500mg DM)), gas production rate (ml/g DM at 2, 4, 5 and 6 h), gas yield (GY_{24h}, ml/g DM disappeared), and gas production parameters of leaves of four shrub species in the presence of 0, 2, or 5 mg of ZADO^{®1} per ml of caecal buffered liquor after 24h of incubation in NZW rabbits

Species	becies A. saligna			A. halimus		L.	L. leucocephala		E. camaldulensis			SEM	Probability			
(S)							-					_			-	
ZADO®	0mg	2mg	5mg	0mg	2mg	5mg	0mg	2mg	5mg	0mg	2mg	5mg	-	S	Z	S×Z
(Z)																
IVGP ₂₄	33.9 ^b	75.7ª	79.9 ^{a*}	45.3 ^b	73.9 ^a	75.5^{a^*}	30.5 ^b	69.6 ^a	78.2 ^{a**}	36.7 ^b	57.8 ^a	73.4 ^{a*}	2.86	0.08	< 0.01	0.31
Rate of gas production																
RGP _{2h}	10.4	15.1	12.1	8.2	13.3	12.2	10.2	13.9	12.4	9.1	12.7	11.2	3.79	0.87	0.16	0.89
RGP _{4h}	8.8	10.2	14.1	6.6	13.4	13.0	5.7	14.1	15.0	5.6 ^b	14.1 ^a	15.6^{a^*}	1.59	0.94	0.01	0.55
RGP _{5h}	8.7	12.5	13.7	7.8 ^b	14.2^{a}	13.2^{a^*}	5.0^{b}	14.3 ^a	14.0^{a^*}	6.2	6.3	14.0	1.55	0.26	0.01	0.23
RGP _{6h}	8.1	12.2	12.6	7.2 ^b	14.2 ^a	12.4^{a^*}	4.3 ^b	15.2ª	11.9^{a^*}	5.6	6.7	13.2	1.43	0.27	0.01	0.13
GY_{24h}	137 ^b	170^{a}	$173^{a^{*}}$	151 ^b	173 ^a	$174^{a^{*}}$	117 ^b	160^{a}	$166^{a^{***}}$	134 ^b	158 ^a	$168^{a^{**}}$	37.1	0.01	< 0.01	0.14
Gas production parameters ²																
В	58.7 ^b	107.8 ^a	122.2 ^{a**}	83.8 ^b	123.4 ^a	116.6 ^{ab*}	57.1 ^b	1242 ^a	118.6 ^{a*}	55.7°	92.3 ^b	121.9 ^{a**}	6.91	0.13	< 0.01	0.21
k	0.19	0.21	0.20	0.14	0.20	0.22	0.23	0.24	0.22	0.24	0.24	0.22	0.126	0.40	0.46	0.40
L	1.9	2.3	1.9	1.0 ^b	2.0^{a}	2.4^{a^*}	0.7	2.2	1.9	0.9 ^b	2.1 ^a	2.3 ^{a*}	0.32	0.61	0.04	0.55
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¹ZADO[®] is a mixture of cellulases, xylanases, α -amylase and proteases from an anaerobic bacterium. ²'B' is the asymptotic gas production (ml/0.5g DM); 'k' is the rate of gas production (/h); 'L' is the initial delay before gas production begins (h). a, b, and c are superscripts following means within a row and foliage species which indicate differences at *P<0.05, **P<0.01, ***P<0.001

CONCLUSIONS

In Egypt, which is a semi-arid country, the incorporation of shrubs and tree leaves into rabbit diet is restricted. Probiotics could improve the utilization of nutrients of the browse leaves *in vitro* as shown in increasing the caecal gas production and the degradability of dry matter. *In vivo* experiments are required to complete this work using a large scale of animals. Our findings suggest a positive effect of ZADO[®] addition to browse leaves in rabbits. ZADO[®] could have a positive effect on the activity of caecal microorganisms in nutrient digestion and degrading secondary compounds of browse leaves.

REFERENCES

- Beauchemin K.A., Colombatto D., Morgavi D.P., Yang W.Z. 2003. Use of exogenous fibrolytic enzymes to improve feed utilization by ruminants. J. Anim. Sci., 81 (E. Suppl. 2), E37–E47.
- Colombatto D., Moulda F.L., Bhat M.K., Owen E. 2007. Influence of exogenous fibrolytic enzyme level and incubation pH on the *in vitro* ruminal fermentation of alfalfa stems. *Anim. Feed Sci. Technol.*, 137, 150-162.
- Cowan M.M. 1999. Plant products as antimicrobial agents. Clin. Microbiol. Rev., 12, 564-582.

Duncan D.B. 1955. Multiple range and multiple F-test. *Biometrics*, 11, 1-42.

- France J., Dijkstra J., Dhanoa M. S., Lopéz S., Bannink A. 2000. Estimating the extent of degradation of ruminant feeds from a description of their gas production profiles observed *in vitro*: derivation of models and other mathematical considerations. *Brit. J. Nutr.*, *83*, *143-150*.
- Fuller R. 1999. Probiotics for Farm Animals. A Critical Review. G.W. Tannocka (ed.) Horizon Scientific Press, Wymondham, England.
- Gado H. 1997. Effect of enzymatic treatments for poor quality roughages on fiber digestibility and nitrogen metabolism in Baladi goats. *Egyptian Journal of Nutrition and Feeds (special issue)*, 50-56.
- Gado H., Metwally H.M., Soliman H., Basiony A.Z.L., El Galil E.R. 2007. Enzymatic treatments of bagasse by different sources of cellulase anzymes. In: Proc. 11th Conference on Animal Nutrition held in Al-Aqsor-Aswan, Egypt in 2 November, 13-18, vol. 10, 607.
- Guerra N.P., Bernardez P.F., Mendez J., Cachaldora P., Castro L.P. 2007. Production of four potentially probiotic lactic acid bacteria and their evaluation as feed additives for weaned piglets. *Anim. Feed Sci. Technol.*, 134, 89–107.
- Juskiewicza J., Semaskaiteb A., Zdunczyka Z., Wroblewskaa M., Gruzauskasb R., Juskiewicz M. 2007. Minor effect of the dietary combination of probiotic *Pediococcus acidilactici* with fructooligosaccharides or polysaccharidases on beneficial changes in the cecum of rats. *Nutr. Res.*, 27, 133–139.
- Makkar H.P.S., Blümmel M., Becker K. 1995. *In vitro* effects and interactions between tannins and saponins and fate of tannins in the rumen. *J. Sci. Food Agric.*, 69, 481-493.
- Marinho M.C., Lordelo M.M., Cunha L.F., Freire J.P.B. 2007. Microbial activity in the gut of piglets: II. Effect of probiotic and probiotic supplementation. *Livestock Sci.*, 108, 236-239.
- Menke K.H., Raab L., Salewski A., Steingass H., Fritz D., Schneider W. 1979. The estimation of the digestibility and metabolisable energy content of ruminant feeding stuffs from the gas production when they are incubated with rumen liquor *in vitro*. J. Agric. Sci., 93, 217-22.

- Ndlovu L.R., Nherera, F.V. 1997. Chemical composition and relationship to in vitro gas production of Zimbabwean browsable indigenous tree species. *Anim. Feed Sci. Technol.*, 69, 121-129.
- Provenza, F.D. 1995. Post ingestive feedback as an elementary determinant of food selection and intake in ruminants. J. Range Manage, 48, 2–17.

Rubanza C.D.K., Shem M.N., Otsyina R., Ichinohe T., Fujihara T. 2003. Nutritive evaluation of some browse tree legume foliages native to semi-arid area in western Tanzania. *Asian Aust. J. Anim. Sci.*, *16*, *1429–1437*.

SAS 1999. User's Guide. Version 6 (4th ed). SAS Institute, Cary, NC, USA.

Salem A.Z.M. 2005. Impact of season of harvest on *in vitro* gas production and dry matter degradability of *Acacia saligna* leaves with inoculum from three ruminant species. *Anim. Feed Sci. Technol.*, 123-124, 67-79.

Salem A.Z.M., Salem M.Z.M., El-Adawy M.M., Robinson P.H. 2006. Nutritive evaluations of some browse tree foliages during the dry season: Secondary compounds, feed intake and in vivo digestibility in sheep and goats. Anim. Feed Sci. Technol., 127, 251–267.

Salem A.Z., Robinson P.H., El-Adawya M.M., Hassan A.A. 2007. *In vitro* fermentation and microbial protein synthesis of some browse tree leaves with or without addition of polyethylene glycol. *Anim. Feed Sci. Technol.*, *138*, *318–330*.

- Sreekanth P., Narayana K., Shridhar N.B., Bhat A. 2006. Toxicity studies of *Calycopteris floribunda* Lam. in calf, rabbit and rat. J. Ethnopharm, 107, 229–233.
- Steel R., Torrie J. 1980. Principles & Procedures of Statistics. 2nd ed. McGraw-Hill International, New York, NY, USA.
- Van Soest, P.J. 1994. Nutritional Ecology of the Ruminant. Cornell University Press, Ithaca, NY, USA.
- Yoon I.K., Stern M.D. 1995. Influence of direct-fed microbials on ruminal microbial fermentation and performance of ruminants: A review. *Asian-Austral. J. Anim. Sci.*, *8*, 533–555.